

P. tricornutum is able to tolerate a range of extreme environmental conditions

WINNERS VS LOSERS **Integrating phenotypic and** transcriptomic responses of

Rhodomonas marina

UNIVERSITY OF HELSINKI

Studies have reported significant declines of cryptophyte algae – generally associated with low temperatures and high salinities – in all areas of the Baltic Sea⁴

Climate change is predicted to intensify the global water cycle, with salty ocean regions getting saltier compared to the global mean and fresh regions (including parts of the Baltic sea) getting fresher¹. Salinity shifts in marine ecosystems could produce filtering effects, eliminating species with narrow environmental constraints, and potentially disrupting their ecosystem functions². It is imperative we predict the effects of such changes on organisms to respond accordingly. However, development of robust predictions about biological responses to environmental change is held back by lack of knowledge about the capacity of organisms to respond to new regimes by plasticity and evolution³.

microalgae to salinity change

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Incubated mixed microalgal cultures of ten species until they reached stationary phase: Diatoma tenuis, Monoraphidium sp., Levanderina fissa, Synechococcus sp. Diacronema lutheri, Kryptoperidinium foliaceum, Alexandrium ostenfeldii, Skeletonema marinoi, Rhodomonas marina, Phaeodactylum tricornutum

PRIOR STUDY N:P = 10 (a) N:P = 16N:P = 80 N:P = 2 Light 🔸 (b)₁₂₅₀ N:P = 16 N:P = 80 N:P = 2 N:P = 10 it 1000 750-500salinity N:P = 16 N:P = 80 N:P = 2 N:P = 10 (C) Primary production was highly affected under hypoosmotic conditions Difference in taxa (a) ₁₅₀. (g) abundance within mixed N:P 100 cultures point to importance of trait 16 variability for functioning in the microalgal 150 community under 150 -10 000) stressful environmental N:P 100 ⁻¹(x 100) 100 conditions. 80 × For more information about this study, please see 150 -N:P S Sonja Repetti's Master's 100 cells thesis: 2

AIM: to understand how the tradeoff between plastic response to salinity stress and resource acquisition affects microalgal stoichiometry.

POC and

increased

(weaker

response

nutrient

limitation)

under

significantly

with salinity

ratio

C:nutrients



AIM: to characterise the microalgal traits underpinning success and failure in responding to changing salinity conditions

Phaeodactylum tricornutum

(1.1) Growth experiment 3 strains 5 salinities (0, 3.5, 5, 20, 35) 6 replicates 200+ generations (~125 days) Morphological and physiological traits: Chlorophyll a, Growth rate, POC, PON, POP, Nutrient uptake **Molecular:** RNA extraction (DGE analysis)

(1.2) Community growth experiment

(1.3) Bioinformatics: Gene family evolution (diatom phylogeny)

Incorporating understudied strains from the Baltic and

North seas

(2.1) Growth experiment 3 strains 5 salinities (0, 3.5, 5, 20, 35) 6 replicates 200+ generations (~160 days) **Morphological and** physiological traits: Chlorophyll a, Growth rate, POC, PON, POP, Nutrient uptake **Molecular:** RNA extraction (DGE analysis)

Rhodomonas marina

(2.2) **Bioinformatics**: Transcriptome comparison with freshwater Cryptomonas sp. CPCC336

> Please provide feedback on this research plan:



Our project will identify genetic expression changes underpinning key traits implicated in responses of microalgae to longer term changes in salinity, and associated shifts in stoichiometry. This will advance understanding of microalgal responses to environmental change, furthering the integration of plastic and evolutionary responses to better predict changes in functional group distribution in marine ecosystem models and develop robust predictions about biological responses to salinity change.



1 Durack, P.J., Wijffels, S.E., Matear, R.J. 2012. Ocean salinities reveal strong global water cycle intensification during 1950 to 2000. Science 336(6080):455-458.

2 Basset, A., et al. 2013. A unifying approach to understanding transitional waters: fundamental properties emerging from ecotone ecosystems. Estuarine, Coastal and Shelf Science 132:5-16.

3 Collins, S., Boyd, P.W., Doblin, M.A. 2020. Evolution, microbes, and changing ocean conditions. Annual Review of Marine Science 12:181-208. 4 Suikkanen, S., et al. 2013. Climate change and eutrophication induced shifts in northern summer plankton communities. PLoS one 8(6):e66475.